

Small Angle Scattering

# by

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### **This Lecture**

- The size of objects measured with SANS
- Typical applications of SANS
- Instrumental resolution how well can size be determined?
- Scattering length density
- Formal expression for scattering cross section
- Scattering from independent particles
- Radius of gyration and Guinier approximation
- Shapes of composite particles
- Particle correlations
- Contrast variation
- Porod scattering
- Scattering from fractals
- Sample requirements for SANS
- Tools for estimating scattering and interpreting data

# Small Angle Neutron Scattering (SANS) Is Used to Measure Large Objects (~10 nm to ~1 μm)

Recall that :

 $\vec{Q} = \vec{k'} \cdot \vec{k_0} = 2k_0 \sin \theta$  for elastic scattering and that

 $\lambda = 2\pi / k = 2\pi / (Q / 2 \sin \theta) = 4\pi \sin \theta / Q$ so we can rewrite Bragg's law  $\lambda = 2d \sin \theta$  as  $d = 2\pi/Q$  or for small  $\theta$  d  $\approx \lambda/2\theta$ 

i.e. small Q => large length scales

Scattering at small angles probes large length scales

Typical scattering angles for SANS are  $\sim 0.3^{\circ}$  to  $5^{\circ}$ 



## Two Views of the Components of a Typical Reactor-based SANS Diffractometer





Note that SANS, like other diffraction methods, probes material structure in the direction of (vector)  $\overrightarrow{Q}$ 

#### The NIST 30m SANS Instrument Under Construction



### Where Does SANS Fit As a Structural Probe?



- SANS resolves structures on length scales of 1 – 1000 nm
- Neutrons can be used with bulk samples (1-2 mm thick)
- SANS is sensitive to light elements such as H, C & N
- SANS is sensitive to isotopes such as H and D

# **Typical SANS Applications**

- Biology
  - Organization of biomolecular complexes in solution
  - Conformation changes affecting function of proteins, enzymes, protein/DNA complexes, membranes etc
  - Mechanisms and pathways for protein folding and DNA supercoiling

#### Polymers

- Conformation of polymer molecules in solution and in the bulk
- Structure of microphase separated block copolymers
- Factors affecting miscibility of polymer blends
- Chemistry
  - Structure and interactions in colloid suspensions, microemeulsions, surfactant phases etc
  - Mechanisms of molecular self-assembly in solutions

#### **Instrumental Resolution for SANS**

$$Q = \frac{4\pi}{\lambda} \sin \theta \implies \left\langle \frac{\delta Q}{Q^2} \right\rangle = \left\langle \frac{\delta \lambda^2}{\lambda^2} \right\rangle + \left\langle \frac{\cos^2 \theta \cdot \delta \theta^2}{\sin^2 \theta} \right\rangle$$
  
For SANS,  $(\delta \lambda / \lambda)_{rms} \sim 5\%$  and  $\theta$  is small, so  $\left\langle \frac{\delta Q}{Q^2} \right\rangle = 0.0025 + \left\langle \frac{\delta \theta^2}{\theta^2} \right\rangle$ 

For equal source - sample & sample - detector distances of L and equal apertures at source and sample of h,  $\delta\theta_{rms} = \sqrt{5/12}$  h/L.

The smallest value of  $\theta$  is determined by the direct beam size:  $2\theta_{\min} \sim 1.5h/L$ At this value of  $\theta$ , angular resolution dominates and

$$\delta Q_{\rm rms} \sim (\delta \theta_{\rm rms} / \theta_{\rm min}) Q_{\rm min} \sim \delta \theta_{\rm rms} 4\pi / \lambda \sim (2\pi / \lambda) h / L$$
  
The largest charged has the strict  $2\pi / \delta Q_{\rm rms} - 2\pi / \delta Q_{\rm rms}$ 

The largest observable object is  $\sim 2\pi/\delta Q_{\rm rms} \sim \lambda L/h$ . This achieves a maximum of about 5  $\mu$ m at the ILL 40 m SANS instrument using

15 Å neutrons.

Note that at the largest values of  $\theta$ , set by the detector size and distance from the sample, wavelength resolution dominates.

# Scattering Length Density

- Remember  $\frac{d\sigma}{d\Omega} = b_{coh}^2 \left\langle \left| \int d\vec{r} \cdot e^{-i\vec{Q}\cdot\vec{r}} n_{nuc}(\vec{r}) \right|^2 \right\rangle$
- What happens if Q is very small?
  - The phase factor will not change significantly between neighboring atoms
  - We can average the nuclear scattering potential over length scales  $\sim 2\pi/10Q$
  - This average is called the scattering length density and denoted  $\rho(\vec{r})$
- How do we calculate the SLD?
  - Easiest method: go to www.ncnr.nist.gov/resources/sldcalc.html
  - By hand: let us calculate the scattering length density for quartz  $SiO_2$
  - Density is 2.66 gm.cm<sup>-3</sup>; Molecular weight is 60.08 gm. mole<sup>-1</sup>
  - Number of molecules per Å<sup>3</sup> = N =  $10^{-24}(2.66/60.08)$ \*N<sub>avagadro</sub> = 0.0267 molecules per Å<sup>3</sup>
  - SLD=Sb/volume = N( $b_{Si} + 2b_O$ ) = 0.0267(4.15 + 11.6) 10<sup>-5</sup> Å<sup>-2</sup> = 4.21 x10<sup>-6</sup> Å<sup>-2</sup>
- A uniform SLD causes scattering only at Q=0; variations in the SLD cause scattering at finite values of Q

# **SLD** Calculation

- www.ncnr.nist.gov/resources/sldcalc.html
- Need to know chemical formula
   and density

Enter	Compound	C6H12
	Density (g/cm^3)	0.86
Not relevant for SLD	<ul> <li>Wavelength (A)</li> </ul>	6
		Calculate
	Neutron SLD	-3.07E-7 (A^-2)
X-ray values	Cu Ka SLD	8.34E-6 +9.36E-9i (A^
	Mo Ka SLD	8.33E-6 +2.08E-9i (A^
Background	Neutron Inc. XS	5.93; 33.4 (cm^-1)
	Neutron Abs. XS	0.0823 (cm^-1)
Determine best sample thickness	Neutron 1/e length	0.166 (cm)

Note units of the cross section – this is cross section per unit volume of sample

#### SANS Measures Particle Shapes and Inter-particle Correlations

$$\frac{d\sigma}{d\Omega} = \langle b \rangle^{2} \int_{space} d^{3}r \int_{space} d^{3}r' n_{N}(\vec{r}) n_{N}(\vec{r}') e^{i\vec{Q}.(\vec{r}-\vec{r}')}$$

$$= \int_{space} d^{3}R \int_{space} d^{3}R' \langle n_{P}(\vec{R}) n_{P}(\vec{R}') \rangle e^{i\vec{Q}.(\vec{R}-\vec{R}')} \left\langle \left| (\rho - \rho_{0}) \int_{particle} d^{3}x.e^{i\vec{Q}.\vec{x}} \right|^{2} \right\rangle_{orientation}$$

$$\frac{d\sigma}{d\Omega} = (\rho - \rho_{0})^{2} \left| F(\vec{Q}) \right|^{2} V_{p}^{2} N_{P} \int_{space} d^{3}R.G_{P}(\vec{R}).e^{i\vec{Q}.\vec{R}}$$

where  $G_P$  is the particle - particle correlation function (the probability that there is a particle at  $\vec{R}$  if there's one at the origin) and  $|F(\vec{Q})|^2$  is the particle form factor :

$$\left|F(\vec{Q})\right|^{2} = \frac{1}{V_{p}^{2}} \left\langle \left|\int_{particle} d^{3}x.e^{i\vec{Q}.\vec{x}}\right|^{2} \right\rangle_{orientation}$$

These expressions are the same as those for nuclear scattering except for the addition of a form factor that arises because the scattering is no longer from point-like particles

#### **Scattering from Independent Particles**

Scattered intensity per unit volume of sample =  $I(\vec{Q}) = \frac{1}{V} \frac{d\sigma}{d\Omega} = \frac{1}{V} \left\langle \left| \int \rho(\vec{r}) e^{i\vec{Q}\cdot\vec{r}} d\vec{r} \right|^2 \right\rangle$ 

For identical particles

$$I(Q) = \frac{N}{V} (\rho_p - \rho_0)^2 V_p^2 \left\langle \left| \frac{1}{V_p} \int_{particle} e^{i\bar{Q}.\bar{r}} d\bar{r} \right|^2 \right\rangle$$



contrast factor

particle form factor  $\left|F(\vec{Q})\right|^2$ 

Note that  $I(0) = \frac{N}{V} (\rho_p - \rho_0)^2 V_p^2$ 

Particle concentration  $c = NV_p/V$  and particle molecular weight  $M_w = \rho V_p N_A$ where  $\rho$  is the particle mass density and N<sub>A</sub> is Avagadro's number

so  $I(0) = \frac{cM_w}{\rho N_A} (\rho_p - \rho_0)^2$  provides a way to find the particle molecular weight

# **Scattering for Spherical Particles**

The particle form factor  $|F(\vec{Q})|^2 = \left| \int_{V} d\vec{r} e^{i\vec{Q}\cdot\vec{r}} \right|^2$  is determined by the particle shape.

For a sphere of radius R, F(Q) only depends on the magnitude of Q:

$$F_{sphere}(Q) = 3V_0 \left[ \frac{\sin QR - QR \cos QR}{(QR)^3} \right] = \frac{3V_0}{QR} j_1(QR) \rightarrow V_0 \text{ at } Q = 0$$

Thus, as  $Q \rightarrow 0$ , the total scattering from an assembly of uncorrelated spherical particles [i.e. when  $G(\vec{r}) \rightarrow \delta(\vec{r})$ ] is proportional to the square of the particle volume times the number of particles.

For elliptical particles replace R by :  $R \rightarrow (a^2 \sin^2 \vartheta + b^2 \cos^2 \vartheta)^{1/2}$ where  $\vartheta$  is the angle between the major axis (a) and  $\vec{Q}$ 



#### Radius of Gyration Is the Particle "Size" Usually Deduced From SANS Measurements

If we measure  $\vec{r}$  from the centroid of the particle and expand the exponential in the definition of the form factor at small Q :

$$F(Q) = \int_{V} d\vec{r} e^{i\vec{Q}\cdot\vec{r}} \approx V_{0} + i\int_{V} \vec{Q}\cdot\vec{r} d^{3}r - \frac{1}{2}\int_{V} (\vec{Q}\cdot\vec{r})^{2} d^{3}r + \dots$$
$$= V_{0} \left[ 1 - \frac{Q^{2}}{2} \frac{\int_{0}^{\pi} \cos^{2}\theta \sin\theta d\theta}{\int_{0}^{\pi} r^{2} d^{3}r} + \dots \int_{V_{0}}^{\pi} \sin\theta d\theta - \frac{\int_{V_{0}}^{\pi} r^{2} d^{3}r}{\int_{V_{0}}^{\pi} d^{3}r} + \dots \right] = V_{0} \left[ 1 - \frac{Q^{2}r_{g}^{2}}{6} + \dots \right] \approx V_{0} e^{-\frac{Q^{2}r_{g}^{2}}{6}}$$

where  $r_g$  is the radius of gyration is  $r_g = \int_V R^2 d^3 r / \int_V d^3 r$ . It is usually obtained from a fit

to SANS data at low Q (in the so - called Guinier region) or by plotting ln(Intensity) v Q<sup>2</sup>. The slope of the data at the lowest values of Q is  $r_g^2/3$ . It is easily verified that the expression for the form factor of a sphere is a special case of this general result.

## Radii of Gyration for Various Shapes\*



v = 10 v = 10 v = 1 (sphere) 0.5 1 1.5  $QR_g$  $R_g = R \sqrt{\frac{2 + v^2}{5}}$ 

0.6

0.4

0.2

0

0

Guinier Appoximation:

 $I(Q) \cong I(0)e^{-\frac{1}{3}R_0^2Q^2}$ 



Note that Guinier approximation works only when  $QR_q < 1$ 

If particles are not mono-disperse, Rg is weighted towards larger particles

$$R_g^2 = \frac{3}{5} \frac{\left\langle R^8 \right\rangle}{\left\langle R^6 \right\rangle}$$

\*Viewgraph from Charles Glinka

# **Beyond Guinier: Form Factors for Simple Shapes**



### Form Factors (cont'd)

3) Thin Disks





Note that the slope of I(Q) v Q that corresponds to the particle shape occurs over a region of Q bounded by the largest and smallest dimensions of the particles

Note also that at large Q the average slope is -4. This is called the Porod region. We will discuss it later. The slope is a manifestation of a smooth 3-dimesnional surface of the particles

# **Calculating Form Factors**

- www.ncnr.nist.gov/resources/simulator.html
- Note: T(1 mm H<sub>2</sub>O) = 0.5; T(1 mm D<sub>2</sub>O) = 0.9 ds/dW (H<sub>2</sub>O) = 1 cm<sup>-1</sup>; ds/dW (D<sub>2</sub>O) = 0.06 cm<sup>-1</sup>



# What Happens if Particles are Lined Up?

Scattering probes structure in the direction of Q





Couette shear cell

#### Determining Particle Size From Dilute Suspensions

- Particle size is usually deduced from dilute suspensions in which inter-particle correlations are absent
- In practice, instrumental resolution (finite beam coherence) will smear out minima in the form factor
- This effect can be accounted for if the spheres are mono-disperse
- For poly-disperse particles, maximum entropy techniques have been used successfully to obtain the distribution of particles sizes



Fig. 4. Plot of  $\ln I(Q)$  vs Q for 3.98 vol.% monodisperse PMMA-H spheres (core C1) in D<sub>2</sub>O/H<sub>2</sub>O mixtures.

#### **Correlations Can Be Measured in Concentrated Systems**

- A series of experiments in the late 1980's by Hayter et al and Chen et al produced accurate measurements of S(Q) for colloidal and micellar systems
- To a large extent these data could be fit by S(Q) calculated from the mean spherical model using a Yukawa potential to yield surface charge and screening length



Fig. 2. Observed ( $\bullet$ ) and calculated (—) scattered intensity I(Q) as a function of momentum transfer Q for a charged micellar dispersion: 0.03 mol dm<sup>-3</sup> hexadecyltrimethylammonium chloride in D<sub>2</sub>O at 313 K. The functions P(Q) and S(Q) are discussed in the text. (1 barn sterad<sup>-1</sup> = 10<sup>-28</sup> M<sup>2</sup> sterad<sup>-1</sup>).

#### **Polymers Studied with SANS**

For a Gaussian chain  $R_g^2 = N\ell^2 / 6$  where  $\ell$  is the segment length More generally,  $R_g \sim N^{\nu}$  where  $\nu = 3/5$  for a good solvent;  $\nu = 1/2$  for a theta solvent or a melt

- Typically Rg is 5 50 nm for most polymers good for SANS
- In addition to examining chain conformation, SANS has been used to probe thermodynamics (e.g. of blends and block copolymers) and to challenge various theories (e.g. random phase approximation)

#### Verification of of the Gaussian Coil Model for a Polymer Melt

- One of the earliest important results obtained by SANS was the verification of that r<sub>g</sub>~N<sup>-1/2</sup> for polymer chains in a melt
- A better experiment was done
   3 years later using a small amount of H-PMMA in D-PMMA (to avoid the large incoherent background) covering a MW range of 4 decades



Fig. 1. SANS results obtained by Kirste, Kruse & Schelten (1972) for 1.2% deuterated poly(methyl methacrylate) (PMMA) in normal PMMA (mol. wt of 250000) plotted in Ornstein–Zernike form. The solid curve represents a Debye function [equation (1)]. This was one of the first quantitative demonstrations of Gaussian coil behavior for bulk polymers.

# **Probing Chain Conformation in Thin Films**





 $R_g$  in the plane of the film is unchanged down to film thickness of  $R_g/2$ 



Thin films of 25% d-PS & 75% PS spun on to Si wafers. 25 wafers => 10 nm total polymer thickness - 0.2 mg



#### **Contrast & Contrast Matching**



\* Chart courtesy of Rex Hjelm



Both tubes contain borosilicate beads + pyrex fibers + solvent. (A) solvent refractive index matched to pyrex;. (B) solvent index different from both beads and fibers – scattering from fibers dominates

#### **Contrast Variation**



### **Isotopic Contrast for Neutrons**

 Hydrogen Isotope	Scattering Length b (fm)	Nickel Isotope	Scattering Lengths b (fm)
$^{1}\mathrm{H}$	-3.7409 (11)	<sup>58</sup> Ni	15.0 (5)
$^{2}D$	6.674 (6)	<sup>60</sup> Ni	2.8 (1)
 <sup>3</sup> T	4.792 (27)	<sup>61</sup> Ni	7.60 (6)
		<sup>62</sup> Ni	-8.7 (2)
		<sup>64</sup> Ni	-0.38 (7)

#### Using Contrast Variation to Study Compound Particles



$$I_1(Q) = (\rho_1 - \rho_2)^2 F_1^2$$

Examples include nucleosomes (protein/DNA) and ribosomes (poteins/RNA)



$$I_{3}(Q) - \frac{(\rho_{1} - \rho_{0})^{2}}{(\rho_{1} - \rho_{2})^{2}} I_{1}(Q)$$

$$= 2(\rho_{1} - \rho_{0})(\rho_{2} - \rho_{0}) F_{1}$$

$$= 0 \text{ at } Q = \pi/R_{12}$$

$$I_{2}(Q) = (\rho_{2} - \rho_{1})^{2} F_{2}^{2}$$

$$I(Q) = \frac{N}{V} \left\langle \left| \Delta \rho_{1} \int_{V_{1}} e^{i\vec{Q}.\vec{r}} d\vec{r}_{1} + \Delta \rho_{2} \int_{V_{2}} e^{i\vec{Q}.\vec{r}} d\vec{r}_{2} \right| \right\rangle$$

$$I(Q) = \Delta \rho_{1}^{2} \left\langle \left|F_{1}(Q)\right|^{2} \right\rangle + \Delta \rho_{2}^{2} \left\langle \left|F_{2}(Q)\right|^{2} \right\rangle + \Delta \rho_{1}^{2} \left\langle \left|F_{2}(Q)\right|^{2} \right\rangle + \Delta \rho_{2}^{2} \left\langle \left|F_{2}(Q)\right|^{2} \right\rangle + \Delta \rho_{1}^{2} \left\langle \left|F_{2}(Q)\right|^{2} \right\rangle + \Delta \rho_{1}^{2} \left\langle \left|F_{2}(Q)\right|^{2} \right\rangle + \Delta \rho_{2}^{2} \left\langle \left|F_{2}(Q)\right|^{2} \left\langle \left|F_{2}(Q)\right|^{2} \right\rangle + \Delta \rho_{2}^{2} \left\langle \left|F_{2}(Q)\right|^{2} \left\langle \left|F_{2}(Q)\right|^{2} \right\rangle + \Delta \rho_{2}^{2} \left\langle \left|F_{2}(Q)\right|^{2} \right\rangle + \Delta \rho_{2}^{2} \left\langle \left|F_{2}(Q)\right|^{2} \left\langle \left|F_{2}(Q)\right|^{2} \right\rangle + \Delta \rho_{2}^{2} \left\langle \left|F_{2}(Q)\right|^{2} \left\langle \left|F_{2$$

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Viewgraph from Charles Glinka (NIST)

#### SANS Has Been Used to Study Bio-machines

- Capel and Moore (1988) used the fact that prokaryotes can grow when H is replaced by D to produce reconstituted ribosomes with various pairs of proteins (but not rRNA) deuterated
- They made 105 measurements of interprotein distances involving 93 30S protein pairs over a 12 year period. They also measured radii of gyration
- Measurement of inter-protein distances is done by Fourier transforming the form factor to obtain G(R)
- They used these data to solve the ribosomal structure, resolving ambiguities by comparison with electron microscopy



Fig. 4. Comparison of neutron map with a mapping of surfaceexposed antigenic sites of ribosomal proteins of the 30S subunit obtained by immune-electron microscopy (Stoeffler & Stoeffler-Meilicke, 1986).

### **Porod Scattering**

Let us examine the behavior of  $|F(Q)|^2 (QR)^4$  at large values of Q for a spherical particle (i.e. Q >> 1/R where R is the sphere radius)

$$|F(Q)|^{2}(QR)^{4} = 9V^{2} \left[ \frac{\sin QR - QR \cos QR}{(QR)^{3}} \right]^{2} (QR)^{4} = 9V^{2} \left[ \frac{\sin QR}{QR} - \cos QR \right]^{2}$$
  

$$\rightarrow 9V^{2} \cos^{2} QR \text{ as } Q \rightarrow \infty$$
  

$$= 9V^{2} / 2 \text{ on average (the oscillations will be smeared out by resolution)}$$
  
Thus  $|F(Q)|^{2} \rightarrow \frac{9V^{2}}{2(QR)^{4}} = \frac{2\pi A}{Q^{4}}$  where A is the area of the sphere's surface.

This is Porod's law and holds as  $Q \rightarrow \infty$  for any particle shape provided the particle surface is smooth.

Another way to obtain it is to expand  $G(r) = 1 - ar + br^2 + ... [with a = A/(2\pi V)]$  at small r and to evaluate the form factor with this (Debye) form for the correlation function.

#### **Scattering From Fractal Systems**

- Fractals are systems that are "self-similar" under a change of scale I.e. R -> CR
- For a mass fractal the number of particles within a sphere of radius R is proportional to R<sup>D</sup> where D is the fractal dimension

#### Thus

 $4\pi R^{2} dR.G(R) = \text{number of particles between distance R and R + dR} = cR^{D-1} dR$   $\therefore G(R) = (c/4\pi)R^{D-3}$ and  $S(\vec{Q}) = \int d\vec{R}.e^{i\vec{Q}.\vec{R}}G(R) = \frac{2\pi}{Q}\int dR.R.\sin QR.(c/4\pi)R^{D-3}$  $= \frac{c}{2}\frac{1}{Q^{D}}\int dx.x^{D-2}.\sin x = \frac{const}{Q^{D}}$ 

For a surface fractal, one can prove that  $S(Q) \propto \frac{const}{Q^{6-D_s}}$  which reduces to the Porod

form for smooth surfaces of dimension 2.

#### Typical Intensity Plot for SANS From Disordered Systems



ln(Q)

### Sedimentary Rocks Are One of the Most Extensive Fractal Systems\*



1010 Theory: D<sub>s</sub>=2.82, ζ=1.2 μm (cm<sup>-1</sup>) ORNL USAN: Scattering intensity d /d ILL D11 SANS ORNL 30 m SANS 10-2 10-4 10-3 10-5 10-2 10-1 Scattering vector Q (Å-1)

Variation of the average number of SEM features per unit length with feature size. Note the breakdown of fractality ( $D_s=2.8$  to 2.9) for lengths larger than 4 microns

\*A. P. Radlinski (Austr. Geo. Survey)

SANS & USANS data from sedimentary rock showing that the pore-rock interface is a surface fractal ( $D_s=2.82$ ) over 3 orders of magnitude in length scale

# Sample Requirements for SANS

- Monodisperse, non-interacting (i.e. dilute)
- Concentration: 1-5 mg/ml
- Volume: 350-700 μl per sample
- Data collection time: 0.5-6 hrs per sample
- Typical biology experiment: 2-4 days
- Deuterated solvent is highly desirable
- Multiple concentrations are usually necessary.
- Specific deuteration may be necessary.
- Multiple solvents of different deuteration are highly desirable -- contrast variation.

#### References

- Viewgraphs describing the NIST 30-m SANS instrument
  - www.ncnr.nist.gov/programs/sans/tutorials/30mSANS\_desc.pdf
- SANS data can be simulated for various particle shapes using the programs available at:
  - www.ncnr.nist.gov/resources/simulator.html
- To choose instrument parameters for a SANS experiment at NIST go to:
  - www.ncnr.nist.gov/resources/sansplan.html
- A very good set of notes on SANS (the SANS Toolbox)
  - http://www.ncnr.nist.gov/staff/hammouda/the\_SANS\_toolbox.pdf